

## Effect of Ketogenic Diet on Monosodium Glutamate-Induced Uterine Fibroids in Female Wistar Rats

O. Kayode (PhD)<sup>1\*</sup>, A. Kayode (PhD)<sup>2</sup>, I. Mgbojikwe (BSc)<sup>1</sup>, D. Rotimi (BSc)<sup>1</sup>

1.Department of Biochemistry, College of Pure & Applied Sciences, Landmark University, Omu-Aran, Kwara State, Nigeria

2.Department of Biochemistry, School of Basic Medical Sciences, Babcock University, Ilishan-Remo, Ogun State, Nigeria

J Babol Univ Med Sci; 23; 2021; PP: 1-8

Received: Jan 17<sup>th</sup> 2020, Revised: Aug 15<sup>th</sup> 2020, Accepted: Aug 31<sup>st</sup> 2020.

### ABSTRACT

**BACKGROUND AND OBJECTIVE:** Ketogenic diet (KGD) is a low-carbohydrate, high-fat and average protein dietary formulation, which has been reported with the ability to ameliorate several metabolic diseases, especially those under the direct influence of hormonal disruptions. Monosodium glutamate (MSG) had been found to induce uterine fibroids in laboratory animals through alterations to hormones, lipids and oxidative state. The present study was conducted to evaluate the effect of KGD on MSG-induced uterine fibroid.

**METHODS:** In this experimental study twenty-four female Wistar rats were divided into four groups of six. Control group received distilled water while the remaining groups were given 300 mg/kg body weight of MSG once a day for 28 days. Thereafter, the three groups of MSG, MSG + keto group 1 and MSG + keto group 2 received standard rat chow, cabbage-based ketogenic diet and coconut-based ketogenic diet, respectively for 42 days. Estrogen, Follicle Stimulating Hormone (FSH), Luteinizing Hormone (LH), malondialdehyde (MDA), Superoxide Dismutase (SOD), catalase (CAT), and Total Cholesterol (TC) were determined in the blood of animals using standard methods and induction of fibroids was confirmed in the uterus by histomorphological measurements.

**FINDINGS:** Significant elevations ( $p < 0.05$ ) were observed in the levels of estrogen ( $1.80 \pm 0.09$  &  $1.27 \pm 0.12$ ), LH ( $1.04 \pm 0.04$  &  $0.39 \pm 0.01$ ), FSH ( $1.51 \pm 0.04$  &  $0.65 \pm 0.03$ ), TC and MDA in the MSG group compared to control. There were significant decreases ( $p < 0.05$ ) in the activities of CAT and SOD enzymes in the MSG group compared to control. Histological analysis confirmed significant reduction ( $p < 0.05$ ) in leiomyomas of the dietary treatment groups compared to that of MSG.

**CONCLUSION:** The study suggests that cabbage- and coconut-based KGD may control the occurrence and progression of fibroids through reduction of oxidative damage and amelioration of hormonal imbalance induced by MSG.

**KEY WORDS:** *Ketogenic Diet, Monosodium Glutamate, Uterine Fibroid, Sex Hormones, Oxidative Damage.*

### Please cite this article as follows:

Kayode O, Kayode A, Mgbojikwe I, Rotimi D. Effect of Ketogenic Diet on Monosodium Glutamate-Induced Uterine Fibroids in Female Wistar Rats. J Babol Univ Med Sci. 2021; 23: 1-8.

\*Corresponding Author: O. Kayode (PhD)

Address: Department of Biochemistry, College of Pure & Applied Sciences, Landmark University, Omu-Aran, Kwara State, Nigeria

Tel: +234 803 7802662

E-mail: kayode.omowumi@lmu.edu.ng

## Introduction

**M**onosodium glutamate (MSG) is the salt of the non-essential amino acid glutamate, which increases appetite by stimulating the appetite center in the hypothalamus. There are certain reports indicating that MSG is toxic to both experimental animals and human beings through induction of lipid dysfunction, testicular dysfunction, obesity and uterine fibroid among others (1-5). MSG induces uterine fibroid in rats by increasing the level of estrogen, cholesterol and total protein (6). MSG is also implicated in the induction and progression of oxidative stress in experimental animals (2).

Oxidative stress describes a condition of imbalance between oxidative radicals and antioxidant defense system (superoxide dismutase, catalase, glutathione, etc.) such that the former is grossly inadequate to combat the radicals, thus leading to the onset of the condition. Antioxidant enzymes play an important role in limiting cellular stress. Superoxide dismutase scavenges  $O_2$  radical by converting superoxide to  $H_2O_2$  and molecular oxygen while catalase brings about the reduction of  $H_2O_2$  and protects tissues from highly reactive hydroxyl radicals (7).

Oxidative stress has been linked to almost all metabolic diseases and has the ability of initiating tissue and eventually organ dysfunction (7). Oxidative stress also enhances progression of tumor growth either malignant or benign such as leiomyomas (8, 9). Uterine fibroids are benign tumors, or leiomyomas of the smooth muscle compartment of the uterus (10-12). Uterine fibroid occurs in most women of reproductive age (13). The symptoms include heavy and irregular bleeding, pelvic pain and pressure, bowel and bladder dysfunction, early pregnancy loss and preterm labor (14). Uterine fibroids are likely to occur in 80% of women by the age of 50 (15). Only 20% to 50% of women are symptomatic although most cases are detected incidentally on imaging in asymptomatic women (15). The pathogenesis is multifactorial, and includes sex hormones such as progestogens and estrogens that proliferate tumor growth, as well as oxidative stress, genetic factors and cytokines (9).

Ketogenic diet (KGD) is a high-fat, low-carbohydrate diet, with enough protein content, which makes the body utilize fat, rather than carbohydrate, as a preferred energy source (16). When a diet rich in carbohydrate is ingested, the substrate generates glucose, which produces ATP for all the organs of the body, including the brain. Ketogenesis is activated in the liver whenever there is a reduction in carbohydrate intake; this catabolizes fat and makes fatty acids and

ketone bodies (17). These ketone bodies are able to cross the blood-brain barrier and provide energy to the brain. Other organ systems also use ketones as an efficient energy source (17). KGD is used as a therapy for weight loss and metabolic function improvement, such as management option for epilepsy, convulsion, reproductive dysfunction, diabetes, heart diseases and tumor growth (1, 18-22).

The treatment of uterine fibroid is classically done through surgery; however, various medical options are available, which provide symptom control while minimizing risks and complications. A large number of clinical trials have evaluated the commonly used medical treatments and potentially effective new ones (23). The two major leading and most promising drugs for uterine fibroids are orally active gonadotropin-releasing hormone receptor (GnRH) blockers and progesterone receptor (PR) modulators (24). Nutrition as a therapy for treatment of toxicity is usually without consequences or adverse side effects and usually cheaper, readily available, and non-invasive. This study explores the option of nutritional control using KGD as an ameliorative tool for monosodium glutamate-induced uterine fibroid in female Wistar rats.

## Methods

This experimental study was approved by the ethics committee of Department of Biochemistry, Landmark University Animal Care Committee with the code of LUAC-0038B.

**Experimental animals:** Twenty-four (24) healthy female Wistar rats weighing  $152 \pm 12$  g were obtained from Biochemistry Animal House, University of Ilorin, Ilorin, Nigeria. The rats were acclimatized for 14 days and during the experiment, they were housed in wooden cages in the animal house of the department of Biochemistry, Landmark University Kwara State, under standard conditions. The animals had access to clean drinking water and rats pellet ad libitum.

**Monosodium Glutamate (MSG) reagent:** MSG was purchased from Sigma Aldrich Chemical Co., St Louis, USA and a stock solution was prepared by dissolving 30 g of MSG in 300 mL of distilled water. With reference to the animals' weights, 300 mg/kg MSG was administered to all the treatment groups once a day for twenty eight days as previously reported (with slight modification) for induction of uterine fibroids in female Wistar rats (25).

**Chemical and Reagents:** Reagent kits used for assay of total cholesterol and triglyceride were products of

Randox Laboratory Limited, UK. All other reagents used were of analytical grade.

**KGD preparation/formulation:** Novel preparation and formulations of low-carbohydrate fibers were made from cabbage and coconut in our laboratory, Department of Biochemistry, Landmark University. Components and ingredients of the diet includes cabbage/coconut (500 g), protein (100 g), fat (250 g), vitamin/minerals (100 g) and food binders (50 g) for every kilogram formulation. The ingredients were thoroughly mixed and water was added to make dough. The dough was rolled, cut into different sizes, and dried in oven at 70 °C for 2 h as described by Kayode et al. (1).

**Animal Treatment and biochemical assays:** The rats were randomly distributed into four groups of six. Control group received distilled water. The monosodium glutamate group (MSG group) received 300 mg/kg of MSG for 28 days. The MSG+keto group 1 was treated with 300 mg/kg body weight of MSG for 28 days followed by cabbage-based ketogenic diet for 42 days while MSG+keto group 2 received 300 mg/kg body weight of MSG for 28 days followed by coconut-based ketogenic diet for 42 days. Treatment was done via oral administration as a single daily dosage. A day after the final exposure, the animals were sacrificed. Blood sample was collected by cardiac puncture into plain sample bottles, and the uterus, kidney, and liver tissues were excised. The serum was prepared by centrifugation at 2500 x g for 15 min and used in determination of hormonal and biochemical assays.

**Hormonal assays:** Oestrogen, Luteinizing Hormone (LH) and Follicle Stimulating Hormone (FSH) levels were determined in the serum based on the principle described by Tietz (26).

**Lipid Assay:** Serum triglyceride was determined according to the principle described by Mochin and Leyva (27). Serum total cholesterol was determined according to the method described by Fredrickson et al. (28).

**Lipid peroxidation and antioxidant enzymes:** The superoxide dismutase (SOD) activity was determined according to the method described by Misra and Fridovich (29). Catalase (CAT) activity was determined according to the method of Singha (30). Malondialdehyde (biomarker for lipid peroxidation) activity was determined according to the method of Satoh (31). Biuret method was used to evaluate the protein concentration of the liver, uterus, kidney and serum as described by Gornall et al. (32).

**Histology:** Histological analysis was carried out to confirm the onset of uterine fibroid in the rats. Uterus tissue of each rat was stained with hematoxylin and eosin (Sigma) and examined under a microscope (Nikon ECLIPSE Ni-U, Tokyo, Japan) at 400× magnification. Images were captured from 10 randomly selected fields per rat, and endothelial thickness was measured using ImageJ software (ImageJ v46a; NIH, U.S.A.).

**Statistical analysis:** Data were expressed as mean±standard error of mean (SEM) and analyzed with one-way ANOVA and student t-test using GraphPad Prism 6 (GraphPad Software Inc., San Diego, California, USA). Tukey's post-hoc test was used to compare mean values and  $p < 0.05$  was considered statistically significant.

## Results

All the treated animals experienced changes in weight during the course of the experiment when compared to the control. The MSG + Keto groups 1 and 2 showed significantly reduced ( $p < 0.05$ ) weight compared to the animals treated with MSG, which had significant weight gain ( $p < 0.05$ ) in comparison with the control (Table 1). The concentration of estrogen was elevated in the animals treated with MSG when compared with the control animals. The estrogen levels were significantly ( $p < 0.05$ ) reduced in those under ketogenic diet, with MSG + Keto group 1 showing greater estrogen level reduction (Table 2). Similar patterns were observed in the levels of follicle stimulating hormone (FSH) and luteinizing hormone (LH) in the experimental groups (Table 2).

The animals treated with MSG showed significantly increased ( $p < 0.05$ ) levels of cholesterol when compared to the control; this was significantly ( $p < 0.05$ ) decreased in the MSG + Keto groups 1 and 2 when compared to the MSG-treated animals (Figure 1). The animals administered with MSG showed significantly ( $p < 0.05$ ) elevated levels of malondialdehyde in the tissues when compared to the control group. These were consequently lowered in the MSG+Keto groups 1 and 2 in the organs when compared to the MSG group (Figure 2).

The animals administered with MSG showed significantly depleted ( $p < 0.05$ ) antioxidant enzymes activities (SOD and CAT) in the tissues when compared to the control. These were significantly ( $p < 0.05$ ) reversed in varying degrees in the MSG + Keto groups 1 and 2 (Figures 3 and 4). The photomicrographs of

control animals shows that the connective tissues and endometrial cells are normal (Figure 5 A). This normal architecture (seen in the control group) has been disrupted in the animals treated with MSG. Severe hyperplasia of the precursor cells was observed in this group (Figure 5 B). However, animals in MSG + Keto

group 1 showed normal outline of the precursor cells and absence of hyperplasia in the endometrium ( Figure 5 C), while animals in MSG + Keto group 2 showed significant reduction of hyperplasia of the precursor cells of the endometrium when compared with the MSG-treated animals (Figure 5 D).

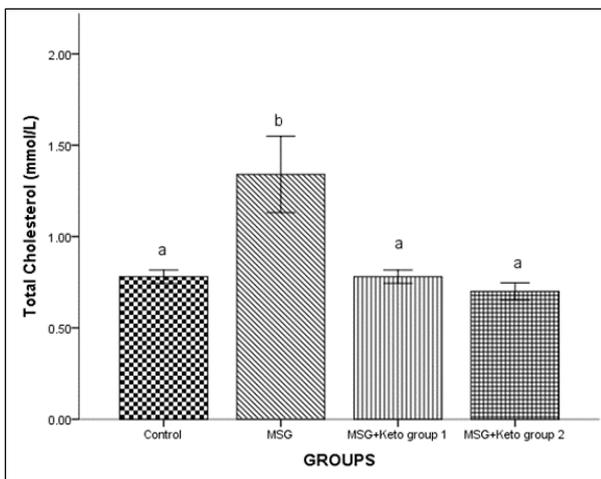
**Table 1. Effect of KGD on the weights of rats with MSG induced fibroids**

Groups	Initial weight (g)	Final weight (g)	Difference in weight (g)	Percentage of weight gain (%)	Percentage of weight loss (%)
Control	175.13±3.5	188.99±3.9	13.86	7.33	-
MSG	161.20±5.5	180.74±6.5	19.5	10.79	-
MSG+keto group 1	147.76±4.8	119.72±7.5	28.04	-	23.42
MSG+Keto group 2	159.79±5.2	144.70±1.5	15.09	-	10.42

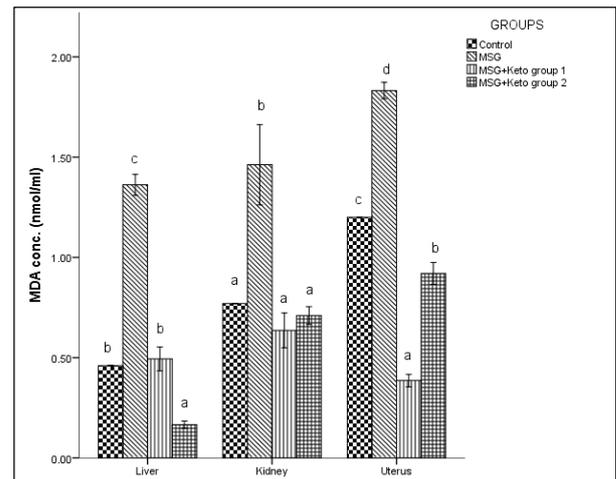
**Table 2. Effect of KGD on Estrogen and gonadotropin concentration in Wistar rats with MSG induced uterine fibroid**

Groups	Estrogen (IU/ml)	FSH (IU/ml)	LH (IU/ml)
Control	1.27±0.12 <sup>b</sup>	0.65±0.03 <sup>a</sup>	0.39±0.01 <sup>a</sup>
MSG	1.80±0.09 <sup>c</sup>	1.51±0.04 <sup>c</sup>	1.04±0.04 <sup>b</sup>
MSG+keto group 1	0.60±0.03 <sup>a</sup>	0.62±0.02 <sup>ab</sup>	0.51±0.01 <sup>a</sup>
MSG+Keto group 2	1.00±0.05 <sup>b</sup>	0.75±0.02 <sup>b</sup>	0.57±0.01 <sup>a</sup>

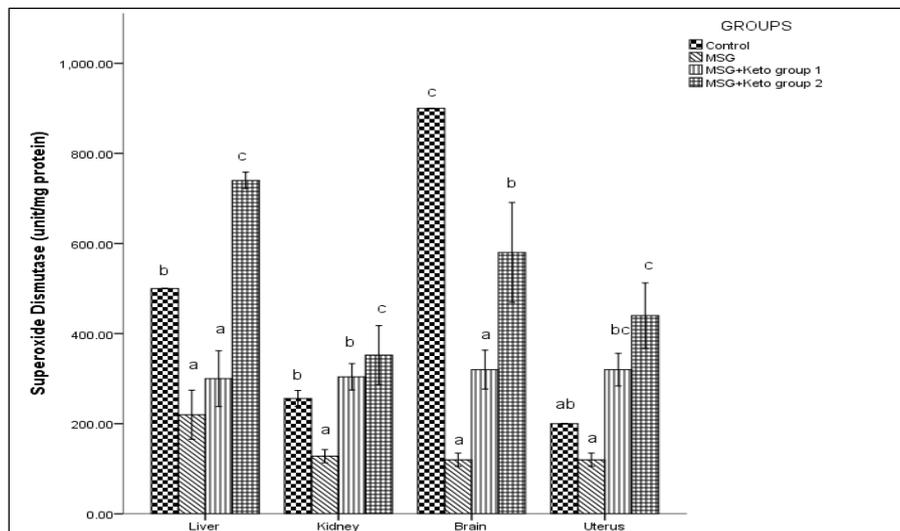
Values are expressed as Mean±SEM. Values bearing different alphabets are significantly different (p≤0.05).



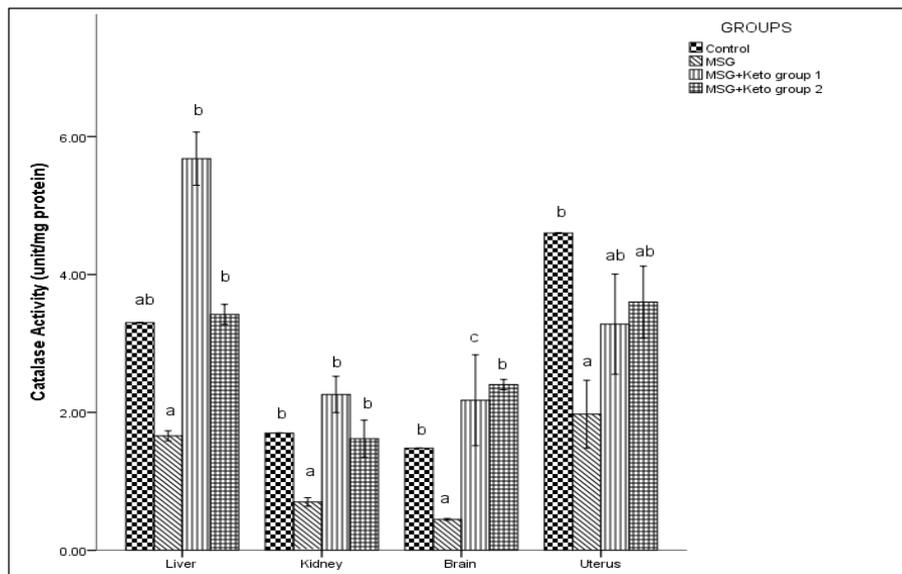
**Figure 1. Effect of KGD on serum total cholesterol in Wistar rats with MSG-induced uterine fibroid.** Values are expressed as Means±SEM. Values with different alphabets are significantly different (p≤0.05).



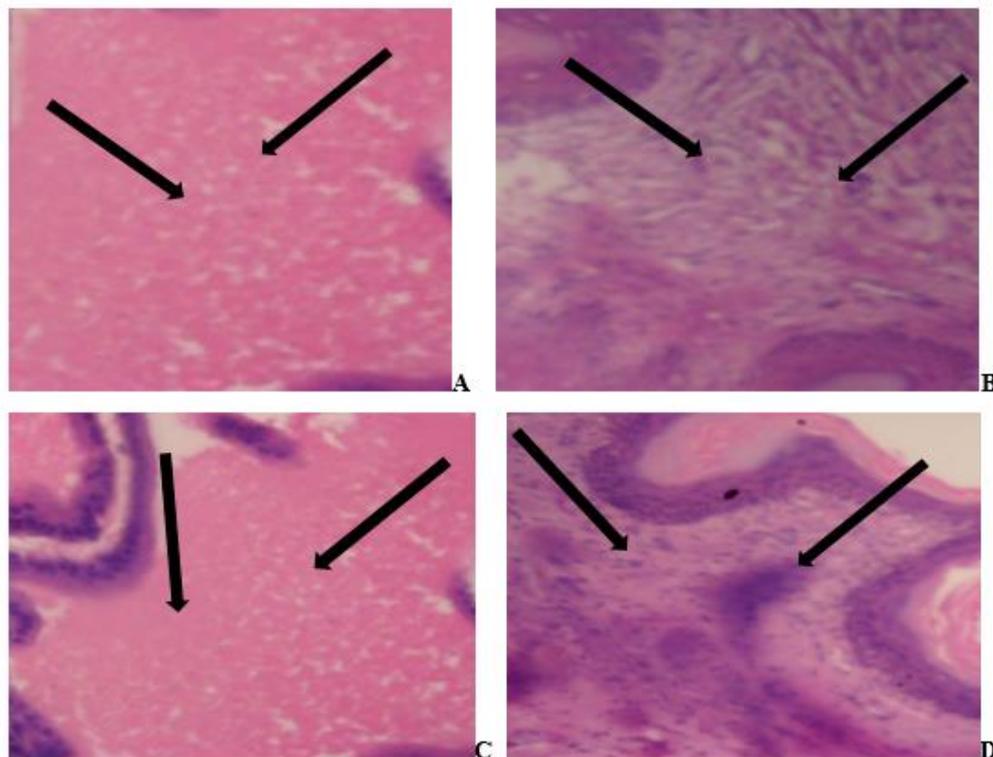
**Figure 2. Effect of KGD on malondialdehyde in Wistar rats with MSG-induced uterine fibroid.** Values are expressed as Means±SEM. Values bearing different alphabets are significantly different (p≤0.05).



**Figure 3. Effect of KGD on SOD activity of kidney of selected tissues in Wistar rats with MSG-induced uterine fibroid.** Values are expressed as Means ± SEM. Values bearing different alphabets are significantly different (p≤0.05).



**Figure 4. Effect of KGD on catalase activity of selected tissues in Wistar rats with MSG induced uterine fibroid.** Values are expressed as Means  $\pm$  SEM. Values bearing different alphabets are significantly different ( $p \leq 0.05$ ).



**Figure 5. Representative photomicrograph of hematoxylin and eosin stained uterus of the control and treated animals (A-D).**

**A:** Photomicrograph of hematoxylin and eosin stained uterus of control animals showing normal connective tissues and endometrial cells outline at X400 magnification

**B:** Photomicrograph of hematoxylin and eosin stained uterus of MSG-treated animals showing severe hyperplasia of the precursor cells (spindle shaped) of the endometrium at X400 magnification

**C:** Photomicrograph of hematoxylin and eosin stained uterus of MSG+KG1-treated animals showing normal outline of the precursor cells (spindle shaped) and absence of hyperplasia in the endometrium at X400 magnification

**D:** Photomicrograph of hematoxylin and eosin stained uterus of MSG+KG2-treated animals showing significant reduction of hyperplasia of the precursor cells (spindle shaped) of the endometrium at X400 magnification

## Discussion

The administration of the two ketogenic diets in this study ameliorated the biochemical alterations associated with MSG-induced uterine fibroids in the Wistar rats. MSG treatment led to a significantly ( $p<0.05$ ) increased weight gain in the animals compared with the control. This finding is in line with previous reports and suggests the ability of chronic consumption of MSG to initiate obesity-related condition in animals and humans (3, 33, 34). Administration of KGD however significantly ( $p<0.05$ ) reduced the weight of the animals (especially those of the cabbage-based formulation), hence supporting previous reports on the use of the diet for achieving effective weight loss in animals (19, 35, 36).

The increase in the level of estrogen in the MSG-treated animals may indicate an increase in the activation of the enzyme aromatase, which catalyzes the conversion of testosterone to estradiol, therefore resulting in elevated estradiol synthesis (37-39). KGD administration was able to ( $p<0.05$ ) reduce the levels of circulating estrogen significantly in the serum using the coconut-based diet, which achieved a higher reduction when compared to the control. Growth of uterine fibroids had been associated with increased levels of estrogen (10, 39). Therefore, the inherent ability of the ketogenic diet to limit the levels of serum estrogen might be one of the mechanisms by which it shrinks the myeloma in the uterus.

The level of FSH and LH, which are precursors directing the production of estrogen in a concentration-dependent manner, was significantly increased ( $p<0.05$ ) in the MSG-treated group probably due to the overproduction of gonadotropin releasing hormone (GnRH), which stimulates the production of LH and FSH from the pituitary gland (40). MSG may therefore direct the onset of hormonal imbalance in females, which often occur prior to the onset of uterine fibroids (41). The KGD ameliorated this increase by significantly reducing ( $p<0.05$ ) the levels of the gonadotropins, which will ultimately contribute to the reduction in estrogen levels.

Obochi et al. (6) reported that MSG administration leads to increase in cholesterol, and estrogen levels, which lead to the induction of fibroid in rats. This is in line with our findings of significant increase ( $p<0.05$ ) in the total cholesterol level for the MSG-treated group when compared to the control group, hence indicating that MSG may cause disorder in lipid metabolism in rats on chronic exposure (25, 42). The elevated serum total

cholesterol was however significantly lowered ( $p<0.05$ ) by the two KGD used in this study. The diet may therefore be useful in managing disease conditions associated with lipid metabolism derangement besides uterine fibroids.

Lipid peroxidation is a marker of oxidative stress (8) and malondialdehyde, which is a by-product of lipid peroxidation, was observed to be elevated in the MSG-treated animals, thereby suggesting that MSG treatment alone may foster the generation of reactive oxygen species (8). Oxidative stress has been implicated previously in uterine fibroids and suggested as one of the mechanisms that is initiated and proliferated (9). KGD administration however caused a significant reduction ( $p<0.05$ ) in MDA levels suggesting its antioxidant and ameliorative potential against MSG-induced fibroids.

Treatment of the animals with MSG brought about a significant decrease ( $p<0.05$ ) in the activities of the measured antioxidant enzymes, hence predisposing the cells to higher probability of oxidative radicalization. The KGD treatment conversely resulted in significant increase ( $p<0.05$ ) in the activity of these enzymes when compared to the MSG-administered group. This observed increase may result in adequate reduction of oxidative stress and effective protection of tissues from highly reactive hydroxyl radicals (43). In addition, MSG-induced uterine hyperplasia was reversed in the groups administered with KGD formulations. The outcome of this work suggest that these ketogenic diet formulations (cabbage- and coconut-based) may possess some bioactive agents that can ameliorate endometrial hyperplasia and also protect against MSG-induced elevated levels of hormones, lipids and oxidative stress that are connected with the initiation and progression of uterine fibroid in rats. A cabbage- and coconut-based KGD may ameliorate the oxidative aberration, hypercholesterolemia and over secretion of female sex hormones induced by MSG intake. The diet may therefore be effective in the management of uterine fibroid caused by repeated oral exposure to monosodium glutamate.

**Conflicts of Interest:** The authors declare no competing financial or non-financial interests.

## Acknowledgment

The authors appreciate the Department of Biochemistry, Landmark University, for providing an enabling environment to carry out this research.

## References

1. Kayode O, Rotimi D, Olaolu T, Adeyemi O. Ketogenic diet improves and restores redox status and biochemical indices in monosodium glutamate-induced rat testicular toxicity. *Biomed and Pharmacol*. 2020;127(2020).
2. Kayode OT, Rotimi DE, Kayode AA, Olaolu TD, Adeyemi OS. Monosodium Glutamate (MSG) Induced Male Reproductive Dysfunction: A Mini Review. *Toxics*. 2020;8(1).
3. Gobatto CA, Mello MA, Souza CT, Ribeiro IA. The monosodium glutamate (MSG) obese rat as a model for the study of exercise in obesity. *Res Commun Mol Pathol Pharmacol*, 2002;111(1-4):89-101.
4. Tawfik MS, Al-Badr N. Adverse Effects of Monosodium glutamate on liver and kidney functions in adult rats and potential protective effect of vitamins C and E. *Food Nutr Sci*. 2012;3:651-9.
5. Zanfirescu, A, Cristea AN, Nitulescu GM, Velescu BS, Gradinaru D. Chronic monosodium glutamate administration induced hyperalgesia in mice. *Nutrients*. 2018;10:1.
6. Obochi GO, Malu SP, Obi-Abang M, Alozie Y, Iyam MA. Effect of garlic extracts on monosodium glutamate (MSG) induced fibroid in Wistar Rats. *Pakistan Journal of Nutrition*. 2009;8(7):970-6.
7. Kayode OT, Kayode AAA, Nwonuma CO. Alcoholic bitters modulates sex hormones and some biochemical parameters of testicular function in male Wistar rats. *F1000Res* 2018;7:1838. Available from: <https://doi.org/10.12688/f1000research.16648.1>
8. Pejic S, Kasapovic J, Todorovic A, Stojiljkovic V, Pajovic SB. Lipid peroxidation and antioxidant status in blood of patients with uterine myoma, endometrial polypus, hyperplastic and malignant endometrium. *Biol Res*. 2017;39:619-29.
9. Markowska A, Mardas M, Gajdzik E, Zagrodski P, Markowska, J. Oxidative stress markers in uterine fibroids tissue in pre and postmenopausal women. *Clin Exp Obstet Gynecol*. 2015;42(6):725-9.
10. Collins S, Arulkumaran S, Hayes K, Jackson S, Impey L. Benign and malignant gynecological conditions. In: Collins, Arulkumaran, Hayes, Jackson, Impey (eds.) *Oxford handbook of obstetrics and gynecology*, 2<sup>nd</sup> ed. Oxford University Press; 2008.p. 641-57.
11. Holdsworth-Carson SJ, Zaitseva M, Vollenhoven BJ, Rogers PA. Clonality of smooth muscle and fibroblast cell populations isolated from human fibroid and myometrial tissues. *Mol Hum Reprod*. 2014;20:250.
12. Wu X, Serna VA, Thomas J, Qiang W, Blumenfeld ML, Kurita T. Subtype-Specific Tumor-Associated Fibroblasts Contribute to the Pathogenesis of Uterine Leiomyoma. *Cancer Res*. 2017;77:6891.
13. Wallach EE, Vlahos NF. Uterine myomas: an overview of development, clinical features, and management. *Obstet Gynecol*. 2004;104:393-406.
14. Bernard NO, Scialli AR, Bobela S. The current use of estrogens for growth suppressant therapy in adolescent girls. *J. Pediat. Adolescent Gynecol*. 2002;15:23-26.
15. Zimmermann A, Bernuit D, Gerlinger C, Schaefer M, Geppert K. Prevalence, symptoms and management of uterine fibroids: an international internet-based survey of 21,746 women. *BMC Women's Health*. 2012;12:6.
16. Kayode, OT, Damilare, ER, Afolayan, OA, Kayode AA. Ketogenic Diet: A Nutritional Remedy for Some Metabolic Disorders. *J Edu, Health Sport*. 2020;10(8).
17. Sanjay K, Rajiv S, Rahul R, Munish D, Deepak, K.; Bharti, K. The Ketogenic Diet. *US Endoc*. 2018;14(2):62-4
18. Hoda EM, Sahar EE, Leila AR, Sally KA. Biochemical effects of a ketogenic diet on the brain of obese adult rats. *J Clin Neur*. 2010;17:899-904.
19. Azar ST, Beydoun HM, Albadri MR. Benefits of ketogenic diet for management of type two diabetes: a review. *J Obes Eat Disord*. 2016;2:1-3.
20. Vargas S, Romance R, Petro JL, Bonilla DA, Galancho I, Espinar S, Kreider RB, Benítez-Porres J. Efficacy of ketogenic diet on body composition during resistance training in trained men: a randomized controlled trial. *J Int Soc Sports Nutr*. 2018;15:31. Available from: <https://doi.org/10.1186/s12970-018-0236-9>
21. Ting R, Dugré N, Allan GM, Lindblad AJ. Ketogenic diet for weight loss. *Canad Fam Phy*. 2018;64.
22. Bolla AM, Caretto A, Laurenzi A, Scavini M, Piemonti L. Low-Carb and ketogenic diets in type 1 and type 2 diabetes. *Nutrients* 2019;11:962.

23. Moroni RM, Vieira CS, Ferriani RA, Candido-dos-Reis FJ, Brito LGO. Pharmacological treatment of uterine fibroids. *Ann Med Health Sci Res.* 2014;4:3.
24. Farris M, Bastianelli C, Rosato E, Brosens I, Benagiano G. Uterine fibroids: an update on current and emerging medical treatment options. *Therap Clin Risk Managt.* 2019;15.
25. Olowofolahan AO, Aina OO, Hassan ET, Olorunsogo OO. Ameliorative potentials of methanol extract and chloroform fraction of *Drymaria cordata* on MSG induced uterine hyperplasia in female Wistar rats. *Eur J Med Plants.* 2017;20(4):1-9.
26. Tietz NW. *Clinical Guide to Laboratory Tests (ELISA)*, 3<sup>rd</sup> ed. W.B. Saunders, Co., Philadelphia: 1995.p. 22-3.
27. Mochin MC, Leyva JA. A new spectrophotometric method for determining triglyceride in serum. *Clin. Chim. Acta,* 1984;142:281-5.
28. Friedewald WT, Levy RI, Fredrickson DS. Estimation of the concentration of low-density lipoprotein cholesterol in plasma, without use of the preparative ultracentrifuge. *Clin Chem.* 1972;18(6):499-502.
29. Xu C, Ghali S, Wang J, Shih DQ, Ortiz C, Mussatto CC, et al. CSA13 inhibits colitis-associated intestinal fibrosis via a formyl peptide receptor like-1 mediated HMG-CoA reductase pathway. *Sci Rep.* 2017;7:16351.
30. Misra HP, Fridovich I. The role of superoxide anion in the autoxidation of epinephrine and a simple assay for superoxide dismutase. *J Biol Chem.* 1972;247(10):3170-5.
31. Beers RF, Sizer IW. A spectrophotometric method for measuring the breakdown of hydrogen peroxide by catalase. *J Biol Chem.* 1952;195(1):133-40.
32. Satoh K. Serum lipid peroxide in cerebrovascular disorders determined by a new colorimetric method. *Clin Chim Acta.* 1978;90(1):37-43.
33. Gornall AG, Badawill CJ, Maxima D. Determination of serum protein by means of the Biuret Reaction. *J Biol Chem.* 1949;177:751-6.
34. He K, Zhao L, Daviglius ML, Dyer AR, Van Horn L, Garside D, Zhu L, Guo D, Wu Y, Zhou B, Stamler J. and for the INTERMAP Cooperative Research Group. Association of monosodium glutamate intake with overweight in Chinese adults: the INTERMAP Study. *Obesity (Silver Spring).* 2008;16(8):1875-80.
35. He K, Du S, Xun P, Sharma S, Wang H, Zhai F, Barry Popkin. Consumption of monosodium glutamate in relation to incidence of overweight in Chinese adults: China Health and Nutrition Survey (CHNS)1-3. *Am J Clin Nutr.* 2011;93:1328-36.
36. Paoli A. Ketogenic diet for obesity: Friend or Foe? *Int. J. Environ. Res. Public Health.* 2014;11:2092-107.
37. Ebbeling CB, Feldman HA, Klein GL, Wong JMW, Bielak L, Steltz SK, et al. Effects of a low carbohydrate diet on energy expenditure during weight loss maintenance: randomized trial. *BMJ.* 2018;363:k4583. Available from: <http://dx.doi.org/10.1136/bmj.k4583>
38. Eweka AO, Eweka A, Om'Iniabohs FAE. Histological studies of the effects of monosodium glutamate of the fallopian tubes of adult female Wistar rats. *N Am J Med Sci.* 2010;2(3):146-9.
39. Zia MS, Qamar K, Hanif R, Khalil M. Effect of monosodium glutamate on the serum estrogen and progesterone levels in female rat and prevention of this effect with diltiazem. *J Ayub Med Coll Abbottabad.* 2014;26(1).
40. Olanrewaju AJ, Olatunji SY, Owolabi JO, Oribamise EI, Omotuyi O I, Desalu ABO. Adeno-Hypophyseal Consequence of Uterine Fibroid and the Effects of Ginger Extract on the Monosodium Glutamate-Induced Tumor. *J Adv Med Medic Res.* 2017;24(11):1-11.
41. Olanrewaju AJ, Owolabi JO, Olatunji SY, Oribamise EI, Omotuyi OI, Desalu ABO. Macroscopic and microscopic molecular-associated treatments of monosodium glutamate-induced uterine fibroid via aqueous extract of ginger rhizomes: A study on adult female Wistar rats. *J Can Tum Int.* 2017;6(4):1-21.
42. Plewka D, MarczyNski J, Morek M, Bogunia E, Plewka A. Receptors of hypothalamic-pituitaryovarian- axis hormone in uterine myomas. *BioMed Res Int.* 2014;521313:13.
43. Olowofolahan AO, Adeosun OA, Afolabi OT, Olorunsogo OO. Effect of methanol extract of *Mangifera indica* on mitochondrial membrane permeability transition pore in normal rat liver and monosodium glutamate-induced liver and uterine damage. *J Compl Alt Med Res.* 2018;5(2):1-14.